REMARKS

Applicants respectfully request reconsideration of the above-identified application in view of the amendments above, and the remarks presented below.

Claims 27-30, 32 and 34 are amended, claims 1-26, 31 and 33 are canceled, and claims 35-45 are added; as a result, claims 27-30, 32, 34 and 35-45 are now pending in this application. The amendments to the claims are made to clarify Applicants' invention, and are not intended to surrender any range of equivalents to which the amended claims may be entitled, such as equivalents of any claim element that are not within the prior art.

At the suggestion of the Examiner, claims 27 and 32 have been amended to recite the full terminology for the acronyms "AAG" and "LPS." The amendments are fully supported by the specification and originally filed claims, and no new matter is added by this amendment.

Support for the amendment to claim 27 is found in the specification at page 1, paragraphs 1-2 and 4.

Support for the amendment to claim 32 is found in the specification at page 9, third paragraph.

Support for the amendment to claim 34 is found in the specification at page 9, sixth paragraph and in claim 26 as originally filed.

Support for new claims 35-37 is found in the specification at page 8, third full paragraph.

Support for new claim 38 is found, for example, in the Abstract of the specification.

Support for new claim 39 is found in the specification at page 10, first full paragraph.

Support for new claims 40-44 is found in the specification at page 9, paragraphs 2-3 and 5.

Support for new claim 45 is found in the specification at page 9, sixth paragraph.

The amendments to the specification address the Examiner's objections at pages 1-3 of the Office Action.

I. §101 Rejection of the Claims

Claims 31-33 were rejected under 35 U.S.C. § 101 because the claimed recitation of a use, without setting forth any steps involved in the process, results in an improper definite of a purpose. Claims 31 and 33 are cancelled, and claim 32 has been amended and no longer recites a "use." Thus, withdrawal of the 35 U.S.C. § 101 rejection of claims 31-33 is proper and respectfully requested.

II. §112 Rejection of the Claims

Claims 27-34 were rejected under 35 U.S.C. § 112, second paragraph, as being indefinite for allegedly failing to particularly point out and distinctly claim the subject matter which Applicant regards as the invention. In particular, the Examiner asserts that the terms "AAG" and "LPS" in claims 27-34 and the phrase "substantially free" in claim 27 are indefinite. In addition, the Examiner asserts that claims 31-33 are indefinite for the recitation of a use without any active, positive steps delimiting how this use is actually practiced. Claims 27-34 are amended to recite the full terminology for "AAG" and "LPS," and claim 27 no longer recites the phrase "substantially free." The cancellation of claims 31 and 33 renders this rejection of claims 31 and 33 moot. Claim 32 is in compliance with 35 U.S.C. § 112, second paragraph.

Therefore, withdrawal of the 35 U.S.C. § 112, second paragraph rejection is respectfully requested.

III. §102 Rejection of the Claims

The Examiner rejected claims 27-34 under 35 U.S.C. § 102(b) as being anticipated by Libert *et al.* (J. Exp. Med., 180, 571-1575 (1994)). The Examiner also rejected claims 27-34 under 35 U.S.C. § 102(b) as being anticipated by Boutten *et al.* (European Journal of Immunology, 22, 2687-295 (1992)). In addition, the Examiner rejected claims 27-34 under 35 U.S.C. § 102(b) as being anticipated by WO 95/07703 (to ALPHA THERAPEUTICS CORP). These rejections are respectfully traversed.

The claims are directed to isolated and purified alpha-1-acid glycoprotein (AAG) having a lipopolysaccharide (LPS) concentration of less than or equal to 0.1 Eu/mg AAG (claims 27-30); a method of treating drug toxicity in a patient in need of such treatment comprising

administering to the patient an effective amount of such AAG (claims 32, 39-45); a pharmaceutical composition comprising such an AAG together with a pharmaceutically acceptable carrier, excipient, or combination thereof (claims 34-37); and to isolated and purified depyrogenated alpha-1-acid glycoprotein (AAG) having a lipopolysaccharide (LPS) concentration of less than or equal to 0.1 Eu/mg AAG (claim 38).

The standard for anticipation is one of strict identity, and to anticipate a claim for a patent a single prior art source must contain all its elements. Hybritech Inc. v. Monoclonal Antibodies, Inc., 231 U.S.P.Q.2d 90 (Fed. Cir. 1986); In re Dillon, 16 U.S.P.Q.2d 1987 (Fed. Cir. 1990). Furthermore, there must be no difference between the claimed invention and the disclosure, as viewed by a person of ordinary skill in the art. Scripps Clinic & Res. Found. v. Genentech, Inc., 18 U.S.P.Q.2d 1001 (Fed. Cir. 1991).

A. Libert *et al.*

The Examiner asserts that Libert *et al.* discloses α-1-acid glycoprotein having over 99% purity, no contaminants, and an endotoxin (*i.e.*, lipopolysaccharide (LPS)) concentration of less than "0.11ng /mg" protein (page 6 of the Office Action). In addition, the Examiner asserts that because Libert *et al.* disclose the *in vivo* administration of AAG, the AAG of Libert *et al.* is depleted of virus. The Examiner also asserts that the recited concentrations of LPS in the claims are "inherent properties" of AAG solutions. Thus, the Examiner alleges that Libert *et al.* discloses "AAG substantially free of LPS" as recited within the limitations of the present claims, thereby anticipating the present claims (page 6 of the Office Action).

Applicants' respectfully note Libert *et al.* disclose that it was their human TNF preparation, not their bAGP preparation, that had an endotoxin level of <0.11 ng/mg protein (see the 13th and 14th lines under the heading "*Reagents*" on page 1571). Libert *et al.* disclose the use of commercial bovine α_1-acid glycoprotein, referred to by Libert *et al.* as "bAGP," which was >99% pure and contained ≤1 ng/mg endotoxin (page 1571, under the heading "*Reagents*"). Libert *et al.* further disclose the *in vivo* administration of the reagent grade bAGP to mice (abstract). Libert *et al.* also disclose that their bAGP preparation had "no contaminating proteins" (see the eighth-ninth lines under the heading "*Reagents*" on page 1571).

The Examiner is respectfully requested to consider that 1 ng/mg endotoxin is equivalent to 1000 pg/mg protein, which is equivalent to 10 Eu/mg endotoxin. In other words, the bAGP of Libert *et al.* contains endotoxin at a concentration that is 100 x greater than the level recited in Applicants' claims. Moreover, the determination by Libert *et al.* as to the absence of "contaminating proteins" in their bAGP preparation was made based upon a denaturing polyacrylamide gel (page 1571). Because LPS is not a protein, it would not appear on such a gel. Moreover, there is nothing in Libert *et al.* that discloses whether or not the bAGP preparation was virus inactivated of virus depleted. Moreover, the mere fact that Libert *et al.* disclose that the bAGP was administered to mice does not refer in any way to the level of endotoxin or virus present in the preparation.

The Examiner is respectfully requested to consider that when "relying upon the theory of inherency, the examiner must provide basis in fact and/or technical reasoning to reasonably support the determination that the allegedly inherent characteristic necessarily flows from the teachings of the applied prior art." M.P.E.P. §2112, citing Ex parte Levy, 17 U.S.P.Q.2d 1461, 1464 (Bd. Pat. App. & Inter. 1990) (emphasis in original). Applicants submit that an assumption has been made that because Libert *et al.* disclose that their bAGP has a purity of greater than 99%, then the bAGP must be "substantially free" of LPS. However, as discussed above, the bAGP of Libert *et al.* contains more than 100 times more endotoxin than the AAG presently claimed. Moreover, there is nothing in Libert *et al.* that discloses removal of endotoxin from the bAGP preparation. Furthermore, as discussed above, the fact that Libert *et al.* report that no "contaminants" were found in their bAGP preparation is irrelevant as to LPS concentration. On the basis that the bAGP was administered to mice, the Examiner concludes that it is must be free of pyrogenic materials (such as LPS) and depleted of viruses. However, the Examiner is urged to consider that the bAGP preparation of Libert *et al.* would inevitably contain LPS, since it was only the present inventors who appreciated the need to remove LPS from AAG in order to make the product therapeutically applicable (see, for example, the present application at page 8, second full paragraph).

Thus, there is nothing in Libert *et al.* that discloses an AAG having the claimed properties, or a method of using such an AAG, or a pharmaceutical composition comprising such

an AAG. Therefore, the pending claims are novel over Libert *et al.* Applicants respectfully request withdrawal of this rejection under 35 U.S.C. § 102 as it relates to Libert *et al.*

B. Boutten *et al.*

The Examiner asserts that Boutten *et al.* disclose a pyrogen-free preparation of AAG that is substantially free of LPS. In particular, the Examiner alleges that passage over a detoxigel column and 0.2μm filter sterilization removed contaminant LPS from Boutten *et al.*'s AAG preparation.

Boutten *et al.* disclose what effect a commercially available preparation of alpha 1-acid glycoprotein (referred to by Boutten *et al.* as "AGP") has on the production of interleukin (IL)-1 beta, IL-6 and tumor necrosis factor (TNF)-alpha by human monocytes, macrophages and the monocytic THP-1 cell line (abstract). Boutten *et al.* disclose that AGP preparations were passed through a Detoxigel column, and then tested for the presence of LPS. Preparations containing more than 200 pg/mg protein were discarded. Thus, the preparations of Boutten *et al.* contained about 200 pg/mg protein. This level is 20 times higher than that claimed in the present application. Moreover, no form of viral inactivation or removal is disclosed by Boutten *et al.*, nor are therapeutic uses of the AGP preparation. The Examiner is also urged to consider that while filter sterilization through a 0.2 μm filter removes bacteria, it does not remove contaminant LPS and/or viruses.

Therefore, the present claims are novel over Boutten *et al.* Applicants respectfully request withdrawal of this rejection under 35 U.S.C. § 102 as it relates to Boutten *et al.*

C. WO 95/07703

The Examiner asserts that WO 95/07703 discloses a pyrogen free AAG having over 99% purity and no contaminants.

WO 95/07703 discloses the purification of α_1-acid glycoprotein (AAG) from Fraction V supernatant (page 2, lines 26-32). However, the disclosed 99% purity of the AAG preparation does not guarantee that the product is endotoxin free or non-pyrogenic. Moreover, although WO 95/07703 refers in a number of places to contaminants, there is no teaching or suggestion

that this is intended to include LPS, and reference to contaminants are to contaminant proteins (not to lipopolysaccharides), as discussed below.

The Examiner is respectfully requested to consider that it is convention to discuss protein purity with regard to the protein composition of the product, while disregarding the presence of non-proteinaceous contaminants. In particular, the figure of 99% purity referred to on page 4 lines 24-25 of WO 95/07703 is in relation to the total *protein* present, and not to other contaminants that may be present. The assay methods referred to in WO95/07703 to test purity (SDS-PAGE, nephelometry and radial immunodiffusion) are used for protein measurement and do not measure the contaminant LPS.

As additional evidence that WO95/07703 does not disclose removal of LPS from the AAG preparation, the Examiner's attention is directed to WO95/22556 (a copy of which is enclosed). WO95/07703 discloses that the AAG in the fraction is bound to an anion exchange medium and then eluted. This occurs because AAG is negatively charged at the pH specified (above 3) because AAG has a pI of 2.7. This fact is disclosed on page 3, third paragraph of WO95/07703. On page 3, WO95/22556 states that a pH level greater than 2 (*i.e.* the conditions under which AAG binds to the anion exchange column) endotoxin aggregates are negatively charged and will bind to positively charged surfaces such as anion exchanges. Therefore, under the conditions specified in the description of WO95/07703, AAG and LPS would both bind to the positively charged anion exchange gel. Thus, this step cannot offer any separation of AAG and LPS - the AAG eluted from the anion exchange column will inevitably contain any LPS present in the original fraction. The subsequent steps referred to on page 4, lines 19-22 of WO95/07703 of diafiltration/ultrafiltration only serves to concentrate AAG and do not remove LPS.

Therefore, the pending claims are not anticipated by WO 95/07703. Applicants respectfully request withdrawal of this rejection under 35 U.S.C. § 102 as it relates to WO 95/07703.

D. The pending claims are not anticipated by the cited art

The Examiner is also urged to consider that LPS is not removed by any standard protein purification step. None of the cited art discloses the removal of LPS to the level as claimed in

the present application. Given that "pure materials are novel *vis-à-vis* less pure or impure materials because there is a difference between pure and impure materials" (M.P.E.P. § 2144.04) and for the reasons discussed herein, withdrawal of the 35 U.S.C. § 102(b) rejection of the claims over Libert *et al.*, Boutten *et al.* and WO 95/07703 is respectfully requested.

Conclusion

Applicants respectfully submit that the claims are in condition for allowance, and notification to that effect is earnestly requested. The Examiner is invited to telephone Applicants' attorney at (612) 371-2106 to facilitate prosecution of this application.

If necessary, please charge any additional fees or credit overpayment to Deposit Account No. 19-0743.

Respectfully submitted,

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(54) Title: PROCESS FOR REMOVING ENDOTOXINS

(57) Abstract

A process for the removal of endotoxins from a biological product, particularly a blood plasma fraction such as albumin, comprises contacting the biological product with a cross-linked hydrophilic matrix comprising a copolymer of allyl dextran and N,N'-methylene bisacrylamide under conditions effective to bind endotoxins in the biological product to the matrix, and recovering purified biological product from which endotoxins have been removed. After recovery of the purified biological product, the matrix can be regenerated under conditions effective to elute bound endotoxins from the matrix.

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PROCESS FOR REMOVING ENDOTOXINS

FIELD OF THE INVENTION

This invention relates to a process for the removal of endotoxins from biologically derived products particularly products such as proteins for therapeutic use. The present invention is particularly but not exclusively directed to the removal of endotoxins from blood plasma fractions such as albumin, as well as biological products derived from gram negative bacterial culture, such as *Escherichia coli* culture.

BACKGROUND OF THE INVENTION

The endotoxins are lipopolysaccharides (LPS) of gram negative bacteria such as *E.coli*, and exist in the outer membrane of the cell envelope. They account for more than half the mass of the outer membrane of the cell envelope and they are constantly shed into the environment of the bacterium (Pearson 1985). The basic unit size of LPS is 10,000 to 20,000. However in aqueous solutions LPS generally exists in vesicles ranging in molecular weight from 300,000 to 1 million (Weary 1985).

The LPS molecule contains 3 distinct chemical regions, the Lipid A region, a central polysaccharide region and the O-antigen region. The Lipid A region resides in the cell membrane when endotoxins are contained within the cell wall. This is linked to a central polysaccharide core and this in turn is linked to the O-antigenic side chain, a repeating oligosaccharide structure which varies with different gram negative species.

The Lipid A region is composed of a glucosamine disaccharide containing phosphate groups and is highly substituted with long chain fatty acids. It is now known that Lipid A is responsible for most, if not all, activity associated with

bacterial endotoxins and that endotoxins must be released from the bacterial surface to be effective (Rietschel and Brade 1992). The biological activities induced by endotoxins are extremely diverse. These are mediated through the activation of macrophages and other cellular components which lead to a wide range of biological effects. In mild doses, endotoxins produce moderate fever and stimulation of the immune system which in turn leads to microbial killing. In higher doses, they produce high fever, hypotension disseminated blood clotting and lethal shock.

The presence of endotoxins in biologically derived products (biologicals) prepared for therapeutic use is of major concern due to the diverse and potentially harmful biological activities of these molecules. Maintaining sterility in processes used in the manufacture of biologicals, together with stringent protocols for the preparation of equipment, helps to ensure products are free of endotoxins. However, raw materials used to manufacture biologicals are often not sterile. Indeed, when the source of a biological is from a gram negative bacterial culture (e.g. a method using an *E.coli* fermentation system to express recombinant protein), the endotoxin levels in the starting material will be very high. In practice, maintenance of sterility throughout an entire process is not always possible or cost effective. Therefore it is often desirable to have methods in place which either destroy or remove endotoxins while maintaining the integrity of the therapeutic biological component.

There have been numerous approaches to achieving destruction or removal of endotoxins (Pearson 1985, Weary 1985). These include hydrolysis with acid or base, oxidation, alkylation, heat treatment and treatment with polymicin B. However with each of these approaches the effect of the inactivation method on the desired biological product must be evaluated.

Furthermore, while pyrogenic activity may be reduced, often endotoxin components remain and the presence of these endotoxin components may be of no benefit in the final product and could possibly be detrimental. It is

therefore preferred to remove these endotoxin components from the final biological product.

Selective binding of endotoxins on charged, hydrophobic or affinity media, or separation on the basis of size can be performed. At pH levels greater than pH2, endotoxin aggregates are negatively charged and will bind to positively charged surfaces such as asbestos or anion exchangers (Weary 1985). Endotoxins will also bind to aliphatic polymers such as polypropylene, polyethylene, polyvinylidene fluoride, polytetrafluoroethylene and hydrophobic chromatographic systems via hydrophobic interactions. Endotoxins can also be specifically removed by affinity chromatography using immobilised polymicin B. Additionally, because endotoxins exist primarily as large molecular weight complexes, they can often be removed from desired components by ultrafiltration or gel filtration methods.

Each of the above mentioned procedures presents a problem. Biological molecules, such as human therapeutic proteins derived from plasma, will in general be positively charged at low pH (i.e. less than pH 4). Although endotoxins are negatively charged at relatively low pH and thus will bind to positively charged resins, many therapeutic proteins are unstable under these conditions. Furthermore, complete resolution between protein and endotoxins cannot always be effected. Hydrophobic chromatographic systems will effectively bind endotoxins but often will also bind the desired biological molecule. Additionally, these hydrophobic chromatographic systems can be difficult to regenerate. Affinity chromatography systems using polymicin B are expensive in terms of media cost. Furthermore, in such systems the support can be difficult to regenerate resulting in a short life for this matrix. Size exclusion chromatography or ultrafiltration can also be used to reduce endotoxin levels. However size exclusion chromatographic systems and ultrafiltration systems will only be useful when there is a substantial size difference between the target biological molecule and the endotoxin molecule. Additionally, size exclusion chromatographic systems suffer from the problem of limited capacity.

The major difficulty in separating endotoxins from proteins lies in designing a support material that exhibits a high specificity for endotoxins but a low specificity for proteins. The ideal support

- should not interact with proteins
- should exhibit a high capacity for endotoxins
- should be able to be regenerated
- should be stable under conditions of operation, including regeneration methods
- should be acceptable for use in the manufacture of therapeutic products.

In work leading to the present invention, it has been found that particular chromatographic gel matrices which have in the past been manufactured and used for gel filtration meet the above criteria, exhibiting a minimal interaction with proteins and a high affinity for endotoxins. Furthermore, it has been established that these matrices are stable under the operational and regeneration systems that have developed for binding of endotoxins and for eluting the bound endotoxin.

SUMMARY OF THE INVENTION

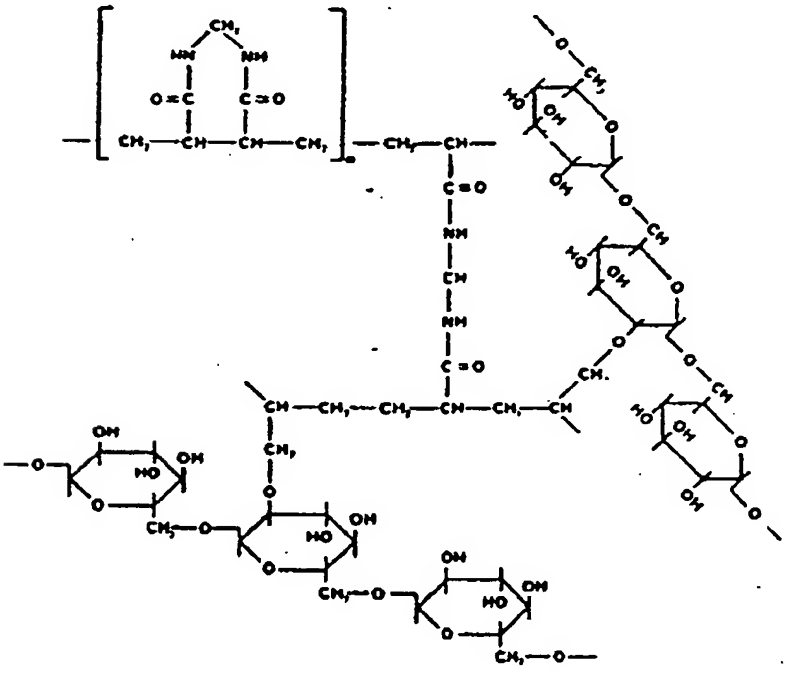
According to the present invention, there is provided a process for the removal of endotoxins from a biological product which comprises contacting said biological product with a cross-linked hydrophilic matrix comprising a copolymer of allyl dextran and N,N'-methylene bisacrylamide under conditions effective to bind endotoxins in said biological product to said matrix, and recovering purified biological product from which endotoxins have been removed.

Preferably, after recovery of the purified biological product, the cross-linked hydrophilic matrix is regenerated under conditions effective to elute bound endotoxins from the matrix.

Throughout this specification, unless the context requires otherwise, the word "comprise", or variations such as "comprises" or "comprising", will be understood to imply the inclusion of a stated integer or group of integers but not the exclusion of any other integer or group of integers.

DETAILED DESCRIPTION OF THE INVENTION

The cross-linked hydrophilic matrix comprising a copolymer of allyl dextran and N,N'-methylene bisacrylamide is preferably a product having the following partial structure:



These products are available commercially as Sephacryl® gel filtration media (Pharmacia, Uppsala, Sweden). The Sephacryl products are sold as gel filtration media for high resolution standard chromatography from millilitre to thousand litre scale. The gel is a cross-linked hydrophilic matrix which at present is sold as six different products, each with a differing porosity range: Sephacryl S200HR, S300HR, S400HR, S500HR and S1000SF. Prior to the present invention, there has been no disclosure of the capacity of the Sephacryl products to bind endotoxins, or of their use in the removal of endotoxins from a biological product.

In a particularly preferred embodiment of the present invention, the cross-linked hydrophilic matrix used for the removal of endotoxins is Sephacryl S200HR.

The biological products which can be purified by removal of endotoxins in accordance with this invention include products for therapeutic use derived from:

(a) plasma (e.g. albumin, immunoglobulins, clotting factors, protease inhibitors and growth factors);

(b) recombinant or cell culture expression systems (e.g. human growth hormone, interferons, cytokines, insulin monoclonal antibodies);

(c) fermentation systems used in the manufacture of vaccines (e.g. components from bordetella pertussis, cultures used in whooping cough vaccine).

In one particularly preferred embodiment, the process of this invention is used for the removal of endotoxins in the production of albumin by fractionation of blood plasma. Methods for separation and purification of albumin and other protein and lipoprotein components of plasma, particularly human plasma, based

on the so-called Cohn fractionation process, have been well known for many years. These methods involve lowering the solubility of protein components of plasma by the addition of ethanol, and fractionation of the plasma under variable conditions of pH, ionic strength, ethanol concentration, protein concentration and temperature. By way of example, one method for the production of albumin using a combination of Cohn fractionation and chromatography is described by Yap *et al.* (1993).

Whilst the plasma fractionation process is carried out under good manufacturing conditions whereby endotoxin contamination of the albumin or other plasma product is generally avoided, accidental contamination can still occur. This often results in the contaminated product being discarded, or alternatively quite expensive purification steps as previously described need to be employed to remove the endotoxins. The present invention provides a process whereby any endotoxin contamination of an albumin or other plasma fractionation can be readily removed, thereby avoiding uneconomical purification steps or even the need to discard the contaminated product. In addition, the process of the present invention can be carried out in the presence of stabilisers such as tryptophan, and sodium octanoate (or sodium caprylate), which are often used in the stabilisation of albumin solutions during pasteurisation.

Studies on the partitioning of endotoxin on Sephacryl S200HR demonstrate that it has a high capacity to bind endotoxins (at least 100 EU/mL of gel). Sephacryl gel matrices have been specifically designed to limit interactions with proteins and for use in the isolation of proteins for therapeutic use, and are designed for use in gel filtration whereby proteins and other biological compounds are separated by size. It has now been established that endotoxins are not resolved on the basis of size using these gel matrices, instead they bind to the matrices and are not eluted under normal aqueous buffer conditions. It has also now been demonstrated that endotoxins can be effectively eluted from Sephacryl gel matrices using 1 M acetic acid, thus allowing the re-use of the matrices. Endotoxins were not eluted from these

matrices using 0.1 M acetic acid or 0.1 M sodium hydroxide, thus indicating that Sephacryl gel matrices will be effective in binding endotoxins over a broad pH range.

The stability of Sephacryl gel matrices has been evaluated under acidic and basic conditions, and results of this study showed that the matrices are stable when exposed to 1 M acetic acid for short time periods (5 days at 37°C). Such stability is more than adequate to allow washing with 1 M acetic acid to elute bound endotoxins in accordance with the present invention, and a long life and repeated use of Sephacryl gel matrices can be expected when used under these conditions.

The endotoxin-binding properties of Sephacryl gel matrices which have now been discovered suggest broad applications for the matrices in the removal of endotoxins from biologically derived products and in particular from blood plasma fractions such as albumin.

Further features of the present invention are more fully described in the following Examples. It is to be understood, however, that this detailed description is included solely for the purposes of exemplifying the present invention, and should not be understood in any way as a restriction on the broad description of the invention as set out above.

BRIEF DESCRIPTION OF THE DRAWING

Figure 1 shows the resolution of endotoxin from albumin monomer by chromatography on Sephacryl S200HR.

Figure 2 shows frontal chromatographic analysis of albumin contaminated with endotoxin.

EXAMPLES

EXAMPLE 1 Endotoxin binding study.

A. **Materials and Methods**

(i) **Endotoxin Concentrate** was prepared from a 5% w/v albumin solution which was allowed to become contaminated with microorganisms by exposure to air for several days at room temperature.

(ii) **Endotoxin assay.** Endotoxin was assayed using a Limulus Amoebocyte Lysate (LAL) test kit (Pyrogent Plus, Bio-Whittaker, Cat. No. N284). Acidic and basic solutions were adjusted to pH 7.0 prior to testing.

(iii) **Sephacryl S200HR (partitioning study):** Sephacryl S200HR (Pharmacia, Uppsala, Sweden) was packed into a column (30 mm diameter x 700 mm) and washed in 100 mM sodium acetate pH 5.5 (2 column volumes) then equilibrated in 50 mM sodium acetate pH 6.8 (4 column volumes). Sample was loaded and eluted in the equilibration buffer (flow rate 2 ml/minute).

(iv) **Albumin solution:** The albumin solution used was a 14% w/v solution in 0.11 M sodium acetate, pH 6.8.

B. **Results**

The sample for Sephacryl S200HR endotoxin partitioning study was prepared by mixing 18 ml of 14% albumin solution with 210 μl endotoxin (450 EU/Ml). A volume of 18 ml was loaded onto the column prepared as described in A(iii). Endotoxin exists generally as high molecular weight complexes (300,000 to 1 million). In this gel filtration system endotoxin was expected to elute in the albumin aggregate, however, no

endotoxin was eluted from this system (Figure 1) indicating that under these chromatographic conditions endotoxin binds to the Sephacryl S200HR column.

EXAMPLE 2

The source material used in this Example was albumin, which had been prepared by Cohn fractionation and formulated as a 5% w/v albumin solution in 140 mM NaCl, 8 mM sodium octanoate, pH 7.0. Sephacryl S200 HR, (Pharmacia) was packed into 3 x 1 ml columns and equilibrated in buffer (50 mM sodium acetate pH 6.8). On each column 2 ml aliquots of a 5% w/v albumin solution that had an endotoxin content of 37.5 to 54 EU/ml was loaded. The column was then washed with 6 column volumes of equilibration buffer to recover residual albumin followed by 6 column volumes of elution buffer, either (i) 0.1 M acetic acid (column 1), (ii) 1 M acetic acid (column 2) or (iii) 0.1 M sodium hydroxide (column 3). The column washing pool and each of the elution peaks were then assayed for endotoxin using a Limulus Amoebocyte Lysate (LAL) test kit (Pyrogen Plus, Bio-Whittaker). The results, showing the efficacy of the three elution buffers for eluting endotoxin bound to the Sephacryl S200HR gel matrix, are summarised in Table 1 below.

Table 1: Efficacy of eluting agent for endotoxins.

Fraction	Total Endotoxin		
	0.1 M sodium hydroxide	0.1 M acetic acid	1 M acetic acid
Initial sample	77 EU	77 EU	108 EU
6 column volume wash (albumin peak)	<0.36 EU recovered	<0.36 EU recovered	<0.36 EU recovered
6 column volume with eluting agent	<0.36 EU recovered	<0.36 EU recovered	95 EU recovered

The results in Table 1 unequivocally demonstrate that Sephacryl S200HR is effective in binding endotoxin. The capacity of the matrix to bind endotoxin is shown here to be greater than 100 EU/ml matrix.

The results of these studies with the three eluting agents showed that only 1 M acetic acid was effective in eluting endotoxin, with essentially all endotoxin that was applied being recovered. Further work was conducted which demonstrated that two column volumes of 1 M acetic acid is sufficient to elute the bulk of bound endotoxin. Washing with 0.1 M sodium hydroxide of 0.1 M acetic acid was ineffective in eluting bound endotoxin.

EXAMPLE 3

The source material used in Examples 3 to 5 was albumin manufactured using a combination of Cohn fractionation and chromatography according to the method of Yap *et al.* (1993). Briefly, the albumin was purified from plasma using Cohn fractionation to separate immunoglobulin from albumin. The crude albumin solution was then purified using ion exchange and gel filtration chromatography. Following ion exchange chromatography, the albumin eluate, in 0.11 M sodium acetate, pH 5.5 was adjusted to pH 6.8 and concentrated to approximately 14% w/v albumin. This material was then processed on Sephacryl S200HR. The albumin monomer was then formulated as 5% w/v albumin, 140 mM NaCl, 8 mM sodium octanoate, pH 7.0.

Sephacryl S200HR (Pharmacia) was packed into a 1 ml column and equilibrated in buffer (50 mM sodium acetate pH 6.8). Varying amounts of a 5% w/v albumin solution that had a high endotoxin content (30-60 EU/ml) was loaded onto the column in sequential runs. After each run, the column was washed with equilibration buffer (6 column volumes) to recover residual albumin and this was pooled with the unbound albumin. Bound endotoxin was subsequently eluted with 1 M acetic acid. The recovered albumin and the 1 M acetic acid eluate were then assayed for endotoxin using a Limulus Amoebocyte Lysate (LAL) test kit (Pyrogent Plus, Bio-Whittaker). The results are summarised in Table 2 below.]

Table 2: Variable loads of endotoxin onto the column.

Load volume (ml)	Endotoxin load EU/ml media	Endotoxin in albumin pool (unbound and residual)	1 M Eluate
0.6	18-36	<0.06 EU/ml	23 EU
2.0	60-120	<0.36 EU/ml	95 EU
3.0	90-180	2.4 EU/ml	46 EU

The results in Table 2 demonstrate that for different loads of endotoxin in albumin, Sephacryl S200HR is effective in removing endotoxin from the albumin and that this endotoxin is subsequently eluted by 1 M acetic acid. After loads of greater than 90 EU/ml, small amounts of endotoxin appear in the albumin pool. The result also demonstrates that the gel can be regenerated as the endotoxin loads were applied sequentially with elution of bound endotoxin after each run.

EXAMPLE 4

The maximum capacity of the Sephacryl gel matrices to adsorb endotoxin was determined using frontal analysis. Approximately 10 ml of a 5% w/v albumin solution that had a high endotoxin content was loaded onto the column, and the unbound material was collected in fractions which were subsequently

assayed for endotoxin using a Limulus Amoebocyte Lysate (LAL) test kit (Pyrogen Plus, Bio-Whittaker). Figure 2 illustrates the unbound albumin passing through the column with no endotoxin present up to 6 ml of loading, after which time endotoxin elutes from the column.

Table 3 shows the equivalent endotoxin loads for this run, for which the maximum capacity appears to be 120 EU/ml media (or 4 ml of 30 EU/ml solution).

Table 3: Frontal analysis data.

Volume loaded	Accumulated endotoxin loaded (IU)	Endotoxin level in unbound fractions
2	>60	<0.06
4	120	0.48
5	150	0.96
5.4	162	31
5.8	180	31
6	186	31
10	310	31

This data demonstrates that endotoxin can be successfully removed from an albumin preparation using Sephacryl S200HR, and that the capacity of this gel matrix to adsorb endotoxins is approximately 120 EU of endotoxin per ml of media.

EXAMPLE 5

Sephacryl S200HR (Pharmacia) was packed into a 1 ml column and equilibrated in an acid buffer, 50 mM sodium acetate pH 5.0. A 3 ml aliquot of a 5% w/v albumin solution that had a high endotoxin content was loaded onto the column, the column was then washed with equilibration buffer to recover residual albumin and this was pooled with the unbound albumin. Bound endotoxin was subsequently eluted with 1 M acetic acid. The recovered albumin

pool and the 1 M acetic acid eluate were then assayed for endotoxin using a Limulus Amoebocyte Lysate (LAL) test kit (Pyrogent Plus, Bio-Whittaker). The results are summarised in Table 4 below.

Table 4: Endotoxin binding under acidic conditions.

Load of endotoxin (EU/ml)	Endotoxin in recovered albumin	Endotoxin in 1 M eluate
>90	<0.06	>30.7

This data demonstrates the ability of the Sephacryl S200HR to adsorb endotoxins from an albumin preparation at low pH.

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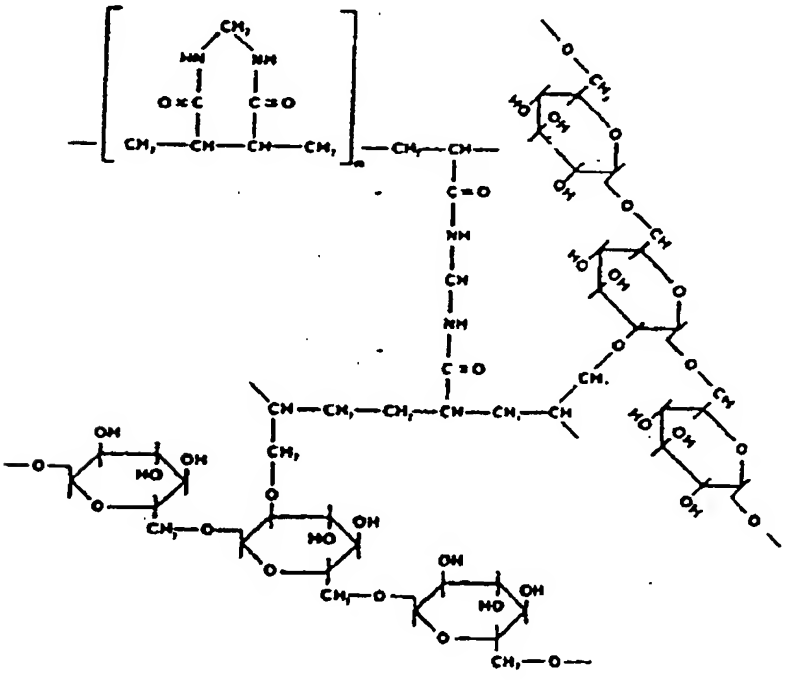
CLAIMS:

1. A process for the removal of endotoxins from a biological product which comprises contacting said biological product with a cross-linked hydrophilic matrix comprising a copolymer of allyl dextran and N,N'-methylene bisacrylamide under conditions effective to bind endotoxins in said biological product to said matrix, and recovering purified biological product from which endotoxins have been removed.

2. A process according to claim 1, wherein the cross-linked hydrophilic matrix comprises a product having the partial structure:



3. A process according to claim 2 wherein the cross-linked hydrophilic matrix is selected from Sephacryl S200HR, Sephacyl S300HR, Sephacryl S400HR and Sephacryl S500HR and Sephacryl S1000SF.

4. A process according to claim 3 wherein the cross-linked hydrophilic matrix is Sephacryl S200HR.

5. A process according to any one of claims 1 to 4, comprising the further step of regenerating the cross-linked hydrophilic matrix under conditions effective to elute bound endotoxins from the matrix.

6. A process according to claim 5 wherein the cross-linked hydrophilic matrix is regenerated by elution with 1 M acetic acid.

7. A process according to any one of claims 1 to 6, wherein the biological product is a protein for therapeutic use.

8. A process according to claim 7, wherein the biological product is a blood plasma fraction.

9. A process according to claim 8, wherein the biological product is an albumin solution.

10. A biological product from which endotoxins have been removed, prepared by the process of any one of claims 1 to 9.

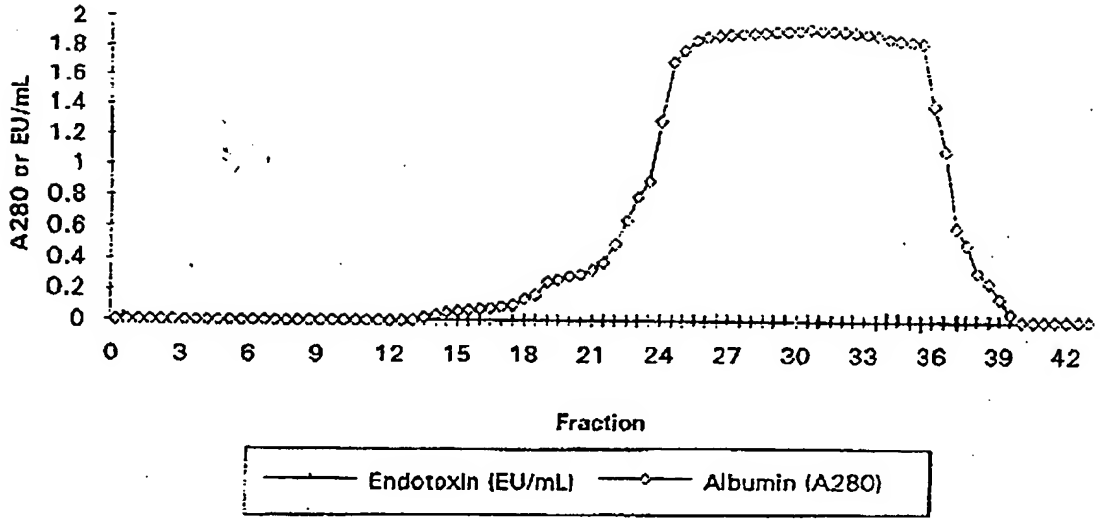


FIGURE 1

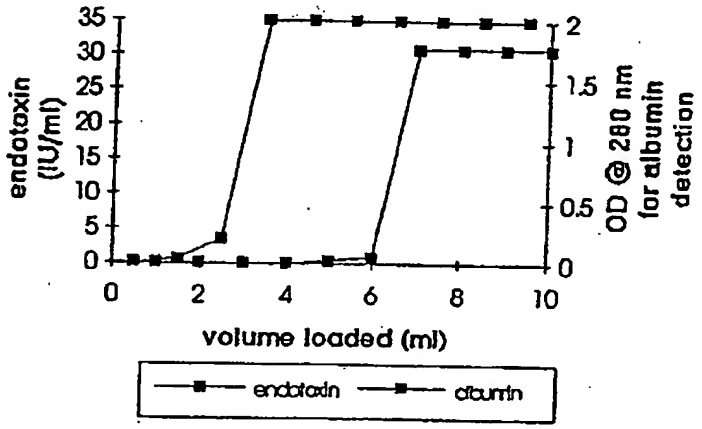


FIGURE 2

INTERNATIONAL SEARCH REPORT

International application No.
PCT/AU 95/00071

A. CLASSIFICATION OF SUBJECT MATTER
Int. Cl.[6] C07K 1/16, C07K 14/245, C07K 14/765

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
FILE WPAT keywords as below.

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched
AU:IPC[6] as above, IPC[5] C07K 3/20; C07K 15/04; C07K 15/06

Electronic data base consulted during the international search (name of data base, and where practicable, search terms used)
FILES WPAT and CASM:keywords allyl dextran; methyl bisacrylamide; sephacryl.

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to Claim No.
Y	US 4086222 (PHARMACIA FINE CHEMICALS AB) 25 April 1978	1-10
Y	JOHANSSON, Bo Lennart and GUSTAVSSON, Jan. Elution behaviour of some proteins on fresh, acid or base treated Sephacryl S-200 HR Journal of Chromatography. 457, 205-13. 1988. See table 1, p 206.	1-10
Y	SARAFIAN, Theodore A; TSAY, Katherine K; FLUHARTY, Arvan L; KIHARA, Hayato. A procedure for pyrogen decontamination of Sephacryl S-300. Biochemical Medicine 28(2) 237-40. 1982	1-10

[X] Further documents are listed in the continuation of Box C.

[X] See patent family annex.

* Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance
"E" earlier document but published on or after the international filing date
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
"O" document referring to an oral disclosure, use, exhibition or other means
"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
30 May 1995	07 JUN 1995 (07.06.95)
Name and mailing address of the ISA/AU AUSTRALIAN INDUSTRIAL PROPERTY ORGANISATION PO BOX 200 WODEN ACT 2606 AUSTRALIA Facsimile No. 06 2853929	Authorized officer ANNE WILCOX Telephone No. (06) 2832243

INTERNATIONAL SEARCH REPORT

International application No.
PCT/AU 95/00071

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate of the relevant passages	Relevant to Claim No.
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Y	AU-B-56812/90 (633925) Centre Regional De Transfusion Sanguine De Lille 13 December 1990	1-10
Y	AU-B-10038/92 (656552) Pasteur Merieux Serums Et Vaccins 13 August 1992	1-10
Y	AU-B-52992/90 (641916) Behringwerke Aktiengesellschaft 11 October 1990	1-10
X	Derwent Abstract Accession No. 90-137268 Class No. A61K 45/02. KR 8902-068-B (Cheil Sugar Co.) 13 January 1987	1-10

This Annex lists the known "A" publication level patent family members relating to the patent documents cited in the above-mentioned international search report. The Australian Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

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	FR B1 2327256	GB A 1554592	JP A2 52047915
	JP B4 56032317	SE A 7511295	SE B 405549
	SE C 405549	US A 4086222	
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	EP A3 367220	US A 5277818	AU A1 43943/93
	AU A1 43943/93	EP A1 643895	US A 5339351
	WO A1 9325036	AU A143943/93	EP A1 643895
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	EP B1 395896	ES T3 2068270	JP A2 2290899
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	US 5345377	WO A19410745	AU A1 52992/93

END OF ANNEX